

Appendix J

Surveillance sampling

The table below provides the timeline and necessary samples to be collected in the three zones.

Surveillance Schedule

	WEEK 1		WEEK 2		WEEK 3		WEEK 4	
	Serum	Swabs	Serum	Swabs	Serum	Swabs	Serum	Swabs
Infected Zone Affected (Minimum 1 mile radius)	YES	YES	NO	YES	NO	YES	NO	YES
Buffer Surveillance Zone (Minimum 1 mile beyond Infected)	YES	YES	NO	NO	NO	NO	NO	YES
Surveillance Zone* (Six miles beyond Control Area)	/	/	/	/	NO	YES	NO	YES

*Premises in the buffer zone will be sampled once beginning two weeks after the first round of sampling begins.

Procedures for Sampling Non-Commercial Premises

Surveillance Team supplies:

- Biosecurity supplies – Tyvek coveralls, rubber boots, boot covers gloves, hair net, respirator mask, disinfectant spray bottle, disinfectant, waterless hand cleaner.
- Brain Heart Infusion broth (BHI) - must be stored in cooler located in “clean” area with ice packs (should be clear amber color, if discolored or cloudy discard).
- Sterile cotton swabs
- Cooler, ice packs or wet ice
- Zip lock bags, large trash bags, duct tape
- GPS unit
- Forms: Avian Influenza Surveillance

Procedures upon arrival at premises:

1. Before exiting the vehicle, each team member should put on one pair of boot covers.

2. Team member will ask the owner to come to the “clean” area or near the vehicle to complete the necessary paperwork. The “clean/dirty” line should be strictly adhered to.
3. Both team member will put on biosecurity equipment (Tyvek, boots, gloves, hair nets, respirator mask) and gather necessary supplies to collect samples. The swabber will duct tape a large plastic bag and two zip lock bags to the Tyvek suit. The necessary number of BHI tubes and swabs will be put into one of the two zip lock bags. The other bag is for the BHI tubes once they are ready for laboratory submission. Both team members will wear two pairs of gloves.

Procedures while on the premises:

1. The Surveillance Team will observe all of the birds to get an accurate inventory of the flock to record on the Avian Influenza Survey Form. They will also determine the health status of the birds.
2. If dead or sick birds are observed, exit the bird housing area immediately and contact a supervisor.
3. Refer to the table to determine which samples need to be collected for each time and zone.

Guidelines for serum sample collections:

1. The catcher will catch and remove birds from cages or use a net to catch loose birds. The owner may also assist with bird handling. Birds will be restrained so that the swabber can collect necessary samples.
2. Blood is collected from the wing vein into a 5 ml red top tube. Collect at least 1 ml of blood from each bird using a sterile blood collection tool (scalpel or needle (22g) and syringe (3cc)).
3. Place tubes in zip lock bag taped to Tyvek suit. Place blood collection instruments in trash bag.

Guidelines for swab sample collections:

1. The catcher will catch and remove birds from cages or use a net to catch loose birds. The owner may also assist with bird handling. Birds will be restrained so that the swabber can collect necessary samples.
2. The swabber will use dry swabs to collect an oropharyngeal swab and cloacal swab from each bird. Collect only cloacal swabs on waterfowl species.
3. Put up to five birds’ samples in each BHI tube, keeping species separate and cloacal and oropharyngeal swabs separate. Write the premises identification number, the species and the location on each tube.
4. Place no more than 5 swabs in a tube. LEAVE SWABS IN TUBE.
5. Dispose of used swabs in trash bag taped to the Tyvek suit of the swabber.

Number of birds to sample per premises or house.

<u>Table 1. Number of birds to sample on each premises.</u>	
<u>Number of birds on the premises</u>	<u>Minimum number of birds to be sampled</u>
<u>10 or less</u>	<u>Sample all</u>
<u>20</u>	<u>15</u>
<u>30</u>	<u>15</u>
<u>40</u>	<u>15</u>
50 or greater	20

Procedures for exiting the premises:

1. Pick up all samples, bag(s) of trash and take to clean/dirty line established earlier.
2. At this area, the swabber will remove all biosecurity clothing using appropriate procedures (see appendix I Biosecurity). The swabber will return to the vehicle to obtain trash bags and zip lock bags in order to double bag samples. At the clean/dirty line the catcher will hold the bags with serum and swab samples so the swabber can spray with Virkon S. The samples will then be placed in the second bag, handed over the line and the second bag will be disinfected with Virkon S. All nets or other equipment should be disinfected at this time.
3. The catcher will remove all Biosecurity clothing using appropriate procedures and cross the clean/dirty line. Double bagged samples will now be placed in a third bag and labeled with the premises identification number, owners name, and species of birds sampled. The third bag is then sprayed with Virkon and placed in the dirty cooler. The trash bag of used biosecurity clothing and trash is sprayed with Virkon and placed in a second bag, which is then sprayed and placed in the dirty area of the vehicle.
4. Both team members will spray their hands with Virkon and then wash their hands with waterless hand cleaner.

Procedures upon returning to the ICP:

1. Place bags containing trash in the designated dumpster.
2. Place ice packs from the dirty cooler, soiled nets, and any other items to be cleaned and disinfected in the designated C&D area.
3. Deliver samples to be transported to the laboratory for testing to the designated sampled receiving and processing area.

4. Vehicles should be washed a commercial car wash that night or in the morning before going out on any other assignments.
5. Team members will shower and change clothes before the next days assignments.

Procedures for transporting samples to the laboratory:

1. A designated courier will transport samples to the laboratory once per day so that samples arrive between 8:00 a.m. and 8:30 a.m.
2. Seal plastic bags to be transported to the laboratory with security tape to ensure integrity of sample tracking and chain of custody.
3. Deliver sealed bags containing samples to laboratory personnel at curbside along with laboratory submission forms.

Procedures for Sampling Commercial premises

Guidelines for Swab Sampling:

1. The swabber will use (separate) dry swabs to collect an oropharyngeal/tracheal swab and a cloacal swab from each bird.
2. Pooled swabs from oropharyngeal/tracheal samples of 5 birds will be placed in BHI (Brain-Heart Infusion broth) tubes. Pooled swabs from cloacal samples of 5 birds will be placed in separate BHI (Brain-Heart Infusion broth) tubes.
3. Place no more than 5 swabs in a tube and LEAVE SWABS IN THE TUBE.
4. Label each tube as either cloacal (C) or tracheal (T). Number all cloacal tubes in succession. Number all tracheal tubes in succession. Indicate which samples are from which house on submission form. Place submission forms in Ziploc bags and attach to bag containing samples.

Sample Shipment:

Depending upon the necessities as determined by situation and the opinion of the Oregon Department of Agriculture, samples for AI testing may be mailed in or collected by courier according to the protocols below.

Mail in Surveillance Protocol

1. Once samples are taken, follow protocol as in “Procedures for Exiting The Premises” for disinfecting and bagging of samples before placing them in the mailer provided to send samples to OSU-VDL.
2. When a mailer is received for testing at OSU-VDL, they will notify ODA Animal Health Lab the same day so that a replacement mailer, submission forms, mailer bags and swabs can be sent out to the producer the next day.

ODA will seek additional recommendations from the USDA’s Standard Operating Procedures HPAI Task Force Manual and the National Avian Influenza Response Plan, August 2006

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