

# Oregon Harmful Algae Bloom Surveillance (HABS) Program

## Public Health Advisory Guidelines Harmful Algae Blooms in Freshwater Bodies



Public Health Division  
Center for Health Protection  
Environmental Public Health Section

Updated April 2018

# Public Health Advisory Guidelines for Harmful Algae Blooms in Freshwater Bodies

Oregon Health Authority Public Health Division  
Center for Health Protection

## Table of Contents

Introduction .....	3
Background .....	3
CyanoHAB Coordination Process .....	4
Criteria for Issuing a Public Health Advisory .....	5
Criteria for Lifting a Public Health Advisory .....	7
Public Notification Methods .....	9
Program Contact Information.....	9

## Appendices

Appendix A: Rationale for and history of standards to issue and lift recreational public health advisories for cyanoHABs.....	10
Appendix B: Toxigenic cyanobacteria and related toxin information .....	12
Appendix C: Exposure pathways.....	21
Appendix D: References.....	24

## Introduction

Cyanobacteria, also known as blue-green algae, are commonly found in many fresh and saltwater environments around the world. Some cyanobacteria species are referred to as toxigenic because they have the potential to produce toxins that can harm people, pets and wildlife.

Some Oregon water bodies are monitored for cyanobacterial harmful blooms (cyanoHABs). The number of waterbodies monitored is affected by available local, state, and federal resources and the costs associated with sampling. Historically the decision-making process for issuing and lifting health advisories varied according to the managing jurisdiction of a specific water body. In 2009, the Oregon Health Authority Public Health Division (OHA) assumed responsibility for the decision-making process and for issuing and lifting public health advisories when cyanoHABs are detected.

The OHA is working to gain a better understanding about the occurrence of cyanoHABs in Oregon and their impact on human health. Funding for Oregon's Harmful Algae Bloom program was through a five-year federal grant from the U.S. Centers for Disease Control and Prevention (CDC). That grant ceased in September of 2013. Currently program staff implement the highest priority activities such as the issuing and lifting of advisories with no funding.

OHA program objectives:

- Track freshwater cyanoHABs with data provided by partner agencies
- Track cases of human and animal illnesses related to cyanoHABs
- Enter environmental and health data for OHA tracking
- Build capacity of our partners to monitor water bodies in a scientifically sound manner with the goal of protecting public health
- Provide technical assistance to partner agencies to assess health risks associated with algal toxins
- Educate and inform the public regarding health risks due to cyanoHABs

## Background

The advisory process guidelines in this document were developed and are modified based on the most current national data and references, and on monitoring data received from our waterbody partners and stakeholders.

These guidelines are used to educate the public and our partners about how and when OHA issues and lifts public health advisories. Public health advisories help to inform the public of the health risks associated with exposure to potentially toxic cyanobacteria in Oregon's recreational fresh waters.

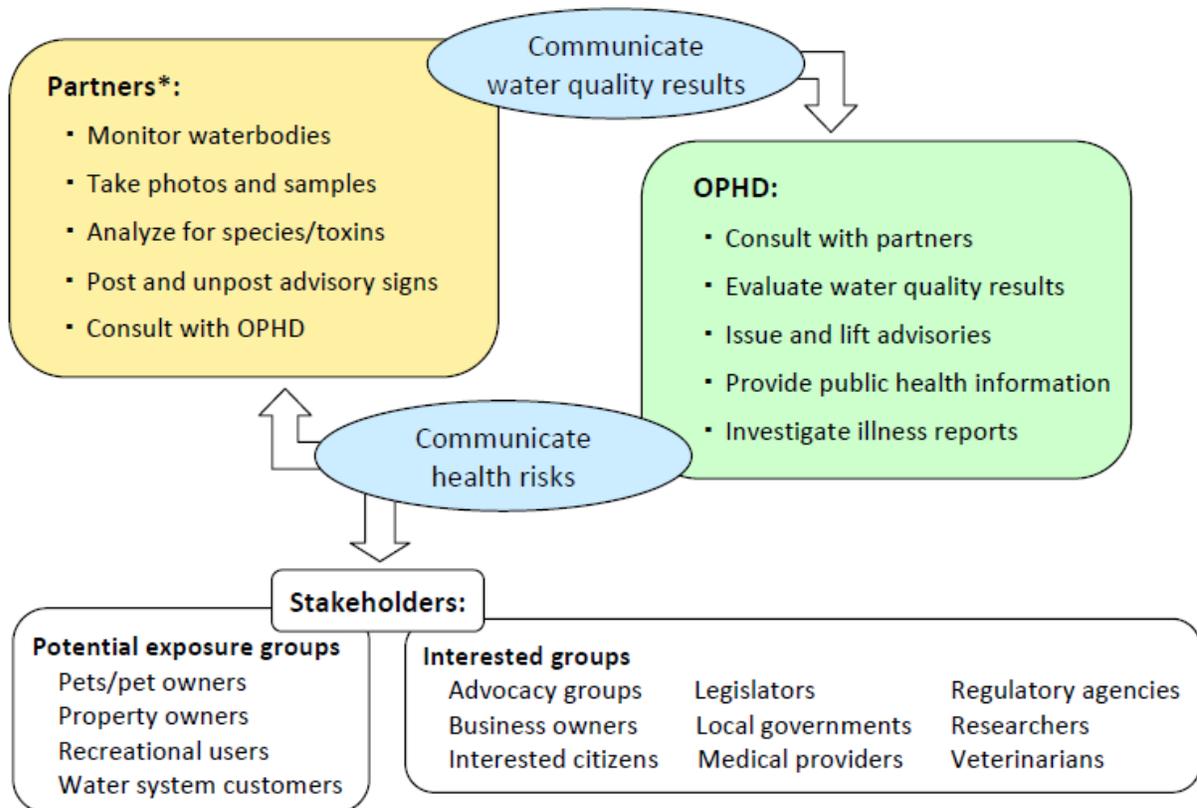
OHA authority for public health and safety fall under Title 36, Oregon Revised Statute (ORS), Chapter 431.035 to 431.530.

## CyanoHAB Coordination Process

Specific actions are involved in monitoring, responding to and communicating information about cyanoHAB blooms.

Coordination among the OHA and its partners and stakeholders is paramount to complete the advisory process from identification and sampling of a bloom to notifying the public of an advisory. Figure 1 depicts the flow of activities among all entities involved in cyanoHAB incidents.

Figure 1. Activities involved in monitoring and responding to cyanoHABs



\*Oregon Department of Environmental Quality, U.S. Forest Service, U.S. Army Corps of Engineers and other waterbody managers.

The main roles of the OHA are to issue and lift health advisories based on water quality data provided by partners and to provide risk communication.

Partners in this effort include the Oregon Department of Environmental Quality, U.S. Forest Service, U.S. Army Corps of Engineers and other waterbody managers.

For the purposes of the OHA public health advisory process, stakeholders are classified in two sub-groups:

- **Exposure:** Those with a greater risk of illness from cyanoHABs through recreational activities. The main routes of exposure are through ingestion and inhalation of contaminated water. Although these toxins are not absorbed through the skin, people with sensitivities can develop a rash when coming into

contact with a cyanoHAB. More information regarding potential routes of exposure is provided in Appendix C.

- Interest: Those with varying levels of need, involvement or interest in program operations or policies are affected by the program, or are intended users of program outcomes/findings.

Table 1.Roles and responsibilities for monitoring and responding to a cyanoHAB

<i>Activity</i>	<i>Lead role</i>	<i>Assist</i>
Monitor	Partners monitor water bodies through on-site observations for evidence of cyanoHABs	OHA provides guidance for establishing a monitoring program
Collect water samples	Partners use scientifically acceptable methods to obtain water samples	OHA provides guidance on sampling techniques
Analyze samples	Partners contract with laboratories that are qualified to perform the required analyses	OHA provides a list of laboratories with appropriate analytic capabilities
Issue or lift advisories	OHA evaluates data and compares test results to established criteria to determine if an advisory should be issued or lifted	OHA informs local health departments before issuing or lifting advisories
Communicate advisory information	OHA informs the general public through advisory news releases, GovDelivery messages, broadcast and print media, an automated electronic list-serv, a toll-free hotline and the HABs website	Partners and local health departments inform constituents of health advisory status through news releases and signage

Ongoing communication between the OHA and partners occurs throughout the bloom season regarding advisory decisions, bloom information, water quality data and illness reports.

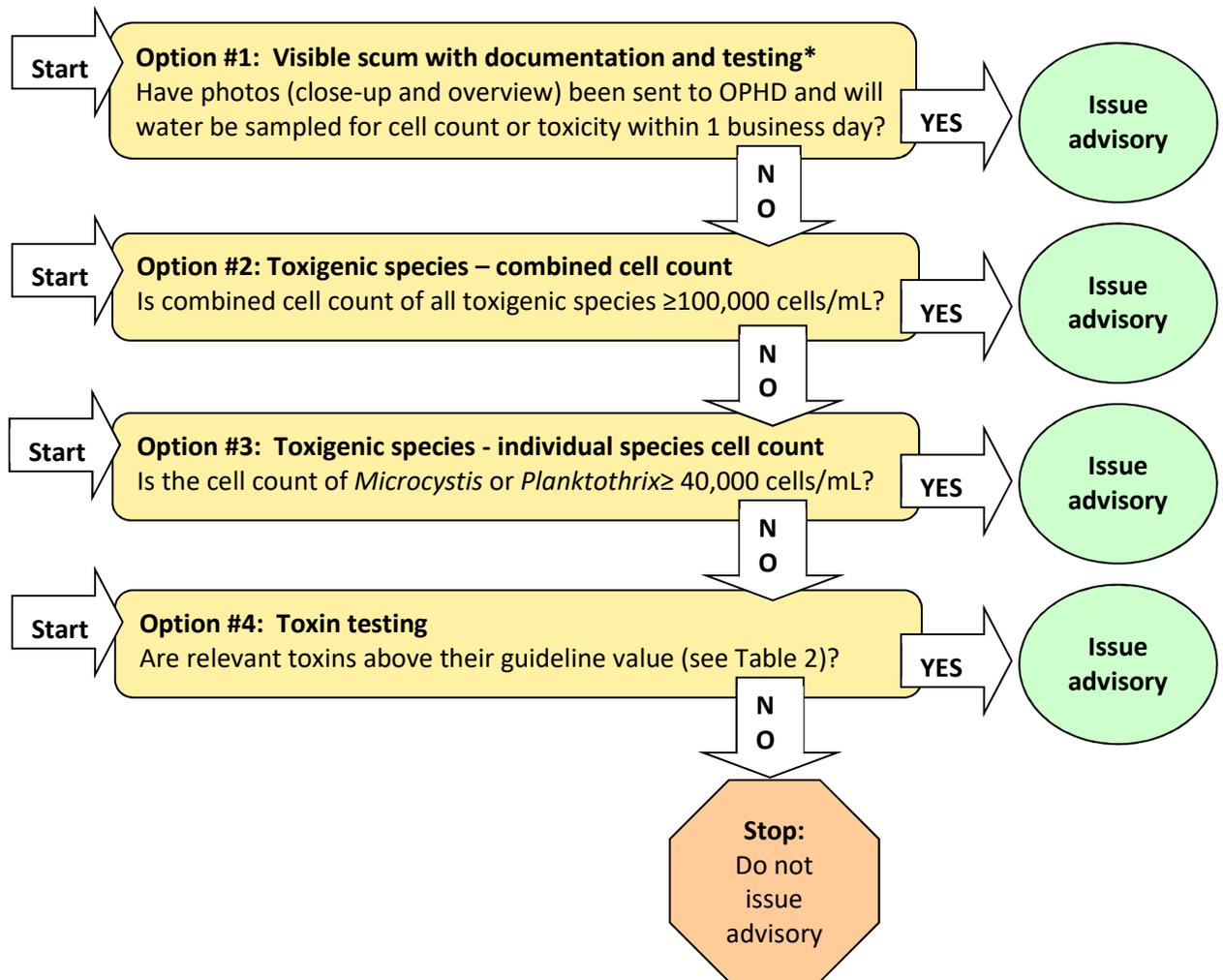
### Criteria for Issuing a Public Health Advisory

OHA is responsible for the decision-making and communication process of issuing and lifting public health advisories. While waiting for the OHA advisory process, local management may post educational signs as a precautionary measure to alert the public of the potential health risks associated with using the water during a cyanoHAB.

OHA criteria for issuing a public health advisory depend on the method selected. Options are:

- Visible scum (with supporting photographs and water analysis)
- Cell counts
- Toxicity levels
- Combinations of two or all three of these options.

Figure 2. OPHD process for issuing public health advisories for a cyanoHAB



Scum is defined as a visible mass of blue-green algae or cyanobacteria identified on the water body. Accumulations of greatest concern are those occurring at or near recreational access points.

OHA guideline values (GVs) for cyanobacterial cell counts are based on the World Health Organization risk categories and research in the field. For cyanotoxins, OHA has updated the GV's for all four cyanotoxins of concern using updated exposure factors based on more current research. More information regarding the rationale used to help determine when advisories should be posted is provided in Appendix A.

#### Additional Guidance on the Toxin Based Monitoring Program: Option 4

Toxin testing provides the most accurate information in terms of protecting public health. Toxin testing also results in health advisory decisions that are based on actual human health risk rather than potential health risk.

Because cyanobacteria often do not produce toxins, even when present in concentrations above OHA's GV's, it is anticipated that Option 4 will result in fewer public health advisories for a given

water body. However, laboratory costs when using Option 4 are higher than those for Options 1 through 3.

OHA’s cyanotoxin GVs, listed in Table 2, are the basis for determining whether an advisory will be issued under Option 4. The OHA Sampling Guidelines document contains detailed information on how to conduct a toxin-based monitoring program.

Table 2. Health advisory GVs for cyanotoxins in Oregon recreational waters (µg/L)

<i>Guideline Value*:</i>	<i>Microcystin</i>	<i>Cylindrospermopsin</i>	<i>Saxitoxin</i>	<i>Anatoxin-a</i>
	4	8	4	8

\*Note that the GVs for all four cyanotoxins have changed from the previous values of 10 and 20 µg/L. See Appendix B for the detailed rationale behind these changes.

OHA has also developed dog-specific GVs. They are for informational purposes only and are not to be used as a basis for issuing public health advisories. These GVs can be found in Appendix C.

Special note: *Aphanizomenon flos-aquae*

*Aphanizomenon flos-aque* (AFA) is a species of cyanobacteria commonly found in Oregon’s fresh waters. Earlier it was determined that AFA either was, or likely was, misidentified in toxicological studies showing toxigenicity. Therefore, AFA was excluded from calculation of combined cell counts of toxigenic species for the purpose of issuing public health advisories.

OHA is in the process of reviewing more current research on AFA to determine whether to reclassify this species as toxigenic in Oregon waters. As before, other species of the genus *Aphanizomenon*, such as *A. gracile* have been demonstrated to produce cyanotoxins. It should be noted that for the 2018 sampling season, AFA will continue to be considered a non-toxigenic species until that review is complete. OHA will update stakeholders and this guidance as needed once the review process is complete. Any changes to the classification of AFA would not be implemented until the 2019 HAB season.

**Criteria for Lifting a Public Health Advisory**

Table 3 summarizes the lifting criteria for advisories issued based on the type of monitoring that led to the advisory.

Table 3. Criteria for lifting advisories

<i>Monitoring option used to generate advisory</i>	<i>Lifting criteria</i>
Option 1: Visible Scum	Initial cell count <u>or</u> toxin results below threshold
Options 2 and 3: Cell counts	Cell counts <u>and</u> toxin results below threshold
Option 4: Toxin based monitoring	Toxin results below threshold

Cyanobacteria can release their toxins during bloom formation and as the bloom is declining, and toxins like microcystin and cylindrospermopsin can take some time to degrade once released. It is possible therefore, to have cell counts below advisory thresholds and still have toxins present. To reduce the risk of exposure to the public from lingering toxins, in all cases, toxin analysis must be completed to lift an advisory.

If an advisory is issued based on Option 1 (visible scum) and initial sample results verify that either cell counts or toxins are below GVs, OHA will immediately lift the advisory. In this case OHA advises continued visual assessment of the bloom and resampling if a change in bloom condition is observed.

If an advisory is issued based on Options 2 or 3 (cell counts above threshold value), OHA will lift advisories when tests show that cell counts **and** toxin concentrations are below the GVs listed in Figure 2 (cell counts) and Table 2 (toxins).

Cell count is required in addition to toxin testing to ensure there is minimal potential for further toxin release. This is because the presence of toxin is what causes illness, while the presence of toxigenic cyanobacteria represents the potential for toxin release. Be sure to choose a laboratory that can analyze for cyanotoxins produced by the cyanobacteria present (see Appendix B, Table B-1).

The accepted method for determining cell counts is Standard Methods Section 10200E and F (also called "SM10200"). We recommend contacting your lab for the most current cost of analyses and for preservation and shipping instructions for your sample.

If an advisory is issued based on Option 4 (toxin results above GVs), OHA will lift the advisory as soon as regular toxin testing indicates that total (intracellular and extracellular) toxin levels are below GVs. In this case, even though the advisory has been lifted, OHA advises continued toxin based monitoring every other week until the bloom is gone to ensure toxin levels remain below GVs. If continued sampling shows an increase in toxins above GVs, a second advisory would be issued.

Commercial laboratories use a variety of comparable methods currently available to analyze for cyanotoxins. When requesting toxin testing, ensure the lab uses a method detection level less than the GVs in Table 2. Note: OHA will not accept field-ready test kits for cyanotoxins as a basis for lifting an advisory. However, these kits may be useful for monitoring the progress of a bloom throughout the season.

Analysis can be costly depending on the method and equipment used. Lab staff can provide you with the most current cost of toxin analyses prior to submitting a sample. In general, the ELISA method is least expensive for determining levels of cyanotoxin in the bloom. ELISA methods are not currently available for anatoxin-a. However, Abraxis has introduced a micro-titer plate format (96T) receptor-binding assay (RBA) kit for anatoxin-a. The kit provides two protocols. The EZ protocol requires no sample preparation and has a range of 5 - 500 ppb. If a lower limit of detection is required, the enhanced sensitivity (ES) SPE sample concentration may be performed. This kit provides a real-time, economical, accurate and sensitive alternative for research and monitoring programs.

Note: All cyanobacteria produce lipopolysaccharides that can cause skin irritation, OHA does not require testing for these endotoxins.

### **Public Notification Methods**

Several concurrent notification methods are used by the OHA in the issuing and lifting of public health advisories. The specific methods are as follows:

*Email:* An email alert is sent to the following groups:

- Health department staff in the county where the waterbody is located
- OHA Partners including agency communications and water quality staff, waterbody managers, watershed council members, basin coordinators, etc.
- Stakeholders including interested citizens, resort owners, advocacy groups, public officials and others. Access to this list is open to all interested Oregonians

*News Releases:* OHA issues statewide news releases which may be picked up and reported by broadcast and print media outlets across Oregon. These releases contain information about the nature and location of the advisory, possible health effects, recommended protective actions and where people can obtain more information. Statewide news releases are also issued when advisories are lifted.

*GovDelivery listserv messages:* A GovDelivery message is sent to notify members about a health advisory issue or lift immediately after the advisory news release is issued. Currently this listserv has over 5,000 members.

*Program Website:* The program maintains a website where advisory information (both issuing and lifting) is immediately posted, providing real-time access to advisory information. Resources for water samplers, prevention tips and general information about cyanoHABs can also be accessed. The website is available at [www.healthoregon.org/hab](http://www.healthoregon.org/hab).

*Hotline:* A statewide toll-free telephone service (877-290-6767) provides updated advisory information to the public, which is particularly helpful for individuals without Internet access.

### **Program Contact Information**

Email: [habhealth@state.or.us](mailto:habhealth@state.or.us)

Phone: (971) 673-0440, Toll Free: (877) 290-6767 and press 4

Website: [www.healthoregon.org/hab](http://www.healthoregon.org/hab)

## **Appendix A: Rationale for and history of standards to issue and lift recreational public health advisories for cyanoHABs**

In 2004 and previous years, lakes were posted when harmful algae cell densities exceeded 15,000 cells/mL. In 2005, a decision was made to no longer use the 15,000 cells/mL threshold as an absolute criterion for posting advisories at recreational access points.

The risk to recreational users at a cell density of 15,000 cells/mL is considered low and includes minor health symptoms such as skin irritation, which are thought to be related to lipopolysaccharide endotoxins found on cell walls. In a study by Pilotto et al, (Pilotto et al., 2004) acute skin irritant effects were tested over a range of cell densities (< 5000 cells/mL to > 200,000 cells/mL) after application of cyanobacterial extracts.

Genera tested included *Dolichospermum* (formerly known as *Anabaena*<sup>1</sup>), *Microcystis*, *Cylindrospermopsis* and *Nodularia*. Approximately 15% of the people reacted to the extracts with mild, self-limiting reactions. Furthermore, no dose-response relationship was established. The absence of a dose-response relationship, and therefore a threshold, makes it difficult to recommend quantitative guidance. Consequently, the focus of advisory postings is on the risk posed by cyanotoxins and the potential for more serious health effects such as nervous system or gastrointestinal disorders.

### Advisory guidelines for algae blooms dominated by *Microcystis* or *Planktothrix*:

A focused risk assessment was conducted to characterize the risk associated with swimming in waters dominated by the cyanobacteria genus *Microcystis* or *Planktothrix*.

According to World Health Organization guidance, 10 µg/L would correspond to approximately 40,000 cells/mL if *Microcystis* were the dominant species (Chorus and Bartram, 1999). *Planktothrix* was included in the additional guidance, since it has the potential to contain higher endocellular microcystin compared with *Microcystis* (Codd et al., 2005). This cell count is associated with a microcystin concentration that is higher than the current guideline value for microcystin of 4 µg/L, but EPA has pointed out that there is uncertainty about the relationship between cell counts and toxin levels, and no other organization has suggested an updated cell count threshold for *Microcystis* or *Planktothrix*. In the absence of better information, OHA will continue to use the 40,000 cells/mL cell count as a threshold.

### Advisory guidelines for algae blooms not dominated by *Microcystis* or *Planktothrix*:

At 100,000 cells/mL, the World Health Organization lists a moderate probability of adverse health effects, based in part on the ability of cyanotoxins to reach levels of concern. As the cell density increases, the potential for frequently occurring cyanobacteria to form scum may increase toxin production by 1000x in a few hours (Chorus and Bartram, 1999).

---

<sup>1</sup>Taxonomy for many types of cyanobacteria is currently being revised. This guidance reflects taxonomy as of 1/2015.

### Rationale for using both cell counts and toxin testing results to lift public health advisories

Several northern California studies conducted between 2005 and 2009 have demonstrated that microcystin concentrations greater than 10 µg/L can be present in rivers and reservoirs where cell counts are below advisory threshold values (Kann and Corum, 2009).

Other research (Manganelli et al., 2010) also suggests that cell count alone is not a good predictor of human health risk. In fact, the State of Washington's Department of Ecology uses only toxin testing data as a basis for public health advisories. The requirement of toxin and cell counts before lifting an advisory is consistent with the OHA goal of public health protection.

Between August 21 and August 30, 2009, four dogs died of acute anatoxin-a poisoning shortly after drinking water from Elk Creek and the Umpqua River, near the confluence of these two streams at Elkton, Oregon.

Water samples collected from the area on September 1, 2009 had no detectable toxigenic cyanobacteria. However, other samples collected from the same areas on the same day revealed detectable levels of anatoxin-a (0.5 µg/L). microcystin was measured at an average concentration of 15 µg/L (1.5 times above the advisory threshold at the time of 10 µg/L). There was no visible bloom or scum reported in that area of the creek when these fatalities occurred.

This case demonstrates that lethal concentrations of cyanobacterial toxin can be present in the absence of detectable toxigenic cyanobacterial cells. This case and other research (Kann and Corum, 2009; Manganelli et al., 2010) demonstrate the importance of measuring both toxin and cell counts before an advisory is lifted.

## Appendix B: Toxigenic cyanobacteria and related toxin information

A variety of species of cyanobacteria are capable of producing toxins that are harmful to people, pets and wildlife (Chorus and Bartram, 1999). The most common toxigenic genera observed during cyanoHABs in Oregon are *Microcystis* and *Dolichospermum*.

*Microcystis* can produce microcystin (liver toxin) and anatoxin-a (neurotoxin). *Dolichospermum*, in addition to producing microcystin and anatoxin-a, can also produce cylindrospermopsin (liver toxin) and saxitoxin (neurotoxin). A complete listing of toxigenic cyanobacteria considered when issuing health advisories in Oregon is presented in Table B-1.

Table B-1. Toxigenic cyanobacteria (data derived from evidence of toxin production (Chorus and Bartram, 1999; Carey et al., 2007; Funari and Testai, 2008; Voloshko et al., 2008))

	Hepatotoxin (liver toxins)			Neurotoxins	
	Microcystin	Nodularin	Cylindrospermopsin	Anatoxin-a	Saxitoxin
<i>Anabaenopsis</i>	+				
<i>Aphanizomenon</i> (Except <i>A. flos-aquae</i> )			+	+	+
<i>Arthrospira</i>	+				
<i>Cyanobium</i>	+				
<i>Cylindrospermopsis</i>			+		+
<i>Dolichospermum</i>	+		+	+	+
<i>Gloeotrichia</i>	+				
<i>Hapalosiphon</i>	+				
<i>Limnothrix</i>	+				
<i>Lyngba</i>					+
<i>Microcystis</i>	+			+	
<i>Nodularia</i>		+			
<i>Nostoc</i>	+				
<i>Oscillatoria</i>	+			+	
<i>Phormidium</i>	+			+	
<i>Planktothrix</i>	+			+	+
<i>Raphidiopsis</i>			+	+	
<i>Schizothrix</i>					
<i>Synechocystis</i>	+				
<i>Umezakia</i>			+		

Note: Table B-1 is at the genus level. Not all species of a given genus produce all the toxins listed for that genus. Once the species involved in a specific bloom have been identified, OHA recommends that waterbody managers contact OHA to determine exactly which toxins could be involved. Taxonomy for many types of cyanobacteria is currently being revised. This guidance reflects taxonomy as of 1/2015.

The primary cyanotoxins of concern in Oregon are microcystin and anatoxin because they have been the toxins most frequently tested and detected. However, cylindrospermopsin has been found above OHA GV and small amounts of saxitoxin have been detected in Oregon. OHA requires

testing for other cyanotoxins listed in Table B-1 to issue and lift advisories when genera reported that produce those toxins are present. Health advisories are not issued solely for algal production of lipopolysaccharides (LPS) as these compounds are produced by most algal species and exposure to LPS compounds typically produce mild, self-limiting rashes in people.

## ***Microcystin***

### Background

Microcystins are the most commonly detected cyanotoxin in the world. Cyanobacteria known to produce Microcystins include *Microcystis*, *Planktothrix*, *Oscillatoria*, *Nostoc*, *Dolichospermum*, *Anabaenopsis* and *Hapalosiphon*. Microcystins are cyclic heptapeptides with about 60 known structural variants (Rinehart et al., 1994). These variations have significant influence on the toxicity and physio-chemical properties of the toxin. The most studied variant is microcystin-LR.

The mechanism of toxicity of microcystins is the inhibition of protein phosphatases which can cause internal hemorrhaging of the liver. While the inhibition of protein phosphatases may be generally cytotoxic, the microcystins primarily target liver cells since they enter cells through a bile acid carrier most abundant on liver cells.

Exposure to microcystin has the potential to cause acute and chronic injury, depending on dose and duration of exposure. Sub-acute damage to the liver is likely to go unnoticed up to levels that are near severe acute damage (Chorus et al., 2000). Two aspects of chronic damage include progressive injury to the liver and tumor-promoting capacity. Microcystins alone have not been classified as carcinogenic. However, microcystins are considered to be tumor promoters based on studies in mice (Falconer and Buckley, 1989).

Most of the mammalian poisonings from the ingestion of microcystin have involved livestock. Symptoms reported from cattle that were exposed to *Microcystis aeruginosa* include generalized weakness, hyperthermia, anorexia, diarrhea, pale mucous membranes, mental derangement, muscle tremors, coma and death within a few days (Short and Edwards, 1990). Symptoms reported from British military recruits exposed to a bloom of *M. aeruginosa* during an exercise included abdominal pain, vomiting, diarrhea, sore throat, blistering of the mouth and pneumonia (Turner et al., 1990).

OHA used a 28-day rat study (Heinze, 1999) as the critical study for determining a tolerable daily intake (TDI). In this study, researchers treated rats with purified microcystin LR in drinking water for 28 days then measured several endpoints. The Heinze study identified a lowest observable adverse effect level (LOAEL) of 50 µg/kg-day.

### Provisional Tolerable Daily Intake

HABS used the LOAEL identified in the Heinze study (Heinze, 1999) described above (50 µg/kg-day) to derive a provisional TDI of 0.05 µg/kg-day as follows:

$$\text{TDI} = \frac{\text{LOAEL}}{\text{UF}}$$

Where:

TDI = Tolerable Daily Intake (0.05 µg/kg-day)

LOAEL = Lowest Observable Adverse Effect Level (50 µg/kg-day)

UF = Uncertainty Factors (1,000 Total = 10 for LOAEL to NOAEL adjustment \*)

10 for interspecies variability \* 10 for individual variability)

This TDI is intended for use with acute or short-term exposure scenarios and may not be protective for chronic or long-term exposures. This recommended TDI should be considered provisional and will be updated to conform to federal guidelines or standards when they are issued, or whenever additional toxicological information becomes available.

Additional support for this TDI: The EPA has used this same TDI as their reference dose (RfD) for microcystins based on currently available research.

#### Provisional Recreational Water Guideline Value

OHA used the TDI of 0.05 µg/kg-day to derive a provisional recreational water **guideline value of 4 µg/L for microcystin:**

$$\text{Guideline Value} = \frac{\text{TDI} \times \text{RSC} \times \text{BW}}{\text{IR}}$$

*Where:*

TDI = Tolerable Daily Intake (0.05 µg/kg-day)

RSC = 0.8 (U.S. EPA 2000a)

BW = Mean body weight of children 6 to < 11 years (31.8 kg) (U.S. EPA 2011)

IR = Recreational water incidental ingestion rate for children (0.33 L/d) at approximately the 90th percentile (U.S. EPA 2011; U.S. EPA 1997)

The TDI was developed by OHA based on oral administration of microcystin-LR via drinking water in rats and effects on the liver (Heinze, 1999).

The mean body weight (BW) of 31.8 kg was used to represent a child between the age of 6 and 11 years. An incidental ingestion rate (IR) was based on EPA guidance for incidental ingestion of recreational water for children at the 90<sup>th</sup> percentile.

The GV for microcystin was the result of new research on exposure factors provided by the Environmental Protection Agency (EPA), specifically affecting body weight and ingestion rate factors.

This recreational water guideline value is based on a provisional TDI. Therefore, this guideline value should also be considered provisional and subject to change should the provisional TDI be updated to accommodate new scientific information.

#### Summary

OHA adopted a health-based GV for microcystin:

- Tolerable Daily Intake: 0.05 µg/kg-day
- **Recreational Water Advisory Guideline Value: 4 µg/L**

The primary limitation in the database relates to chronic toxicity. Because OHA only intends to apply these GVs in acute or short-term exposure scenarios, there is no extrapolation from acute to chronic toxicity. Therefore, OHA considered the uncertainty factor for database limitations to be unnecessary.

## **Anatoxin-a**

### Background

OHA reviewed available literature on the toxicology of anatoxin-a (Astrachan et al., 1980; Astrachan and Archer, 1981; Fawell and James, 1994; Chorus and Bartram, 1999; Fawell et al., 1999b; Duy et al., 2000; Rogers et al., 2005; Codd et al., 2005; Falconer and Humpage, 2005; van Apeldoorn et al., 2007; Burch, 2008; Pegram et al., 2008) as well as accepted and proposed threshold values used in other governmental jurisdictions (New Zealand Ministry of Health, 2002; USEPA, 2006; Washington Department of Health, 2008).

OHA selected a study conducted by Fawell et al. (Fawell and James, 1994; Fawell et al., 1999b) as the critical study for derivation of a TDI. In this study, groups of 10 male and 10 female mice were orally treated with anatoxin-a every day for 28 days at 4 doses (0, 100, 500, and 2,500 µg/kg-day). The mice were observed for health effects over the course of the experiment and many health-related endpoints and physiological parameters were measured (Fawell and James, 1994; Fawell et al., 1999b).

Three animals died during the study. One of the deaths was not related to treatment but rather resulted from animals fighting in their cages. Two of the deaths, one at 500 µg/kg-day and one at 2,500 µg/kg-day, could have been related to treatment. None of the surviving animals had any observable adverse health effects. Therefore, OHA selected 100 µg/kg-day as the no observable adverse effect level (NOAEL).

### Provisional Tolerable Daily Intake

OHA used the NOAEL identified in the Fawell et.al. study (Fawell and James, 1994; Fawell et al., 1999b) described above (100 µg/kg-day) to derive a provisional TDI of 0.1 µg/kg-day as follows:

$$\text{TDI} = \frac{\text{NOAEL}}{\text{UF}}$$

*Where:*

TDI = Tolerable Daily Intake (0.1 µg/kg-day)

NOAEL = No Observable Adverse Effect Level (100 µg/kg-day)

UF = Uncertainty Factors (1,000 Total = 10 for interspecies variability \* 10 for Individual variability \* 10 for limitations in the database)

This TDI is intended only for use in acute or short-term exposure scenarios because the toxicity study upon which this TDI is based was short-term. Because most exposures in Oregon are acute or short-term, an acute or short-term TDI is the most useful.

OHA applied a total uncertainty factor of 1,000. This number is a composite of 3 types of uncertainty about this TDI. First, the critical study was conducted in mice, which may have physiological differences in the way they absorb, distribute, metabolize and excrete anatoxin-a relative to humans. Mice may also be more or less sensitive to anatoxin-a toxicity than humans. Therefore, an uncertainty factor of 10 was applied to account for these potential interspecies differences in sensitivity to anatoxin-a.

Second, humans could have considerable individual variability in their sensitivity to anatoxin-a. For example, a child may be more sensitive than an adult or people with certain genetic traits may be more sensitive than the general population. Therefore, another uncertainty factor of 10 was

applied to account for this individual variability. Finally, OHA applied an additional uncertainty factor of 10 due to limitations in the database. Very few applicable studies have been conducted to identify dose-response relationships to anatoxin-a administered orally. Therefore, this uncertainty factor accounts for the possibility that additional studies in the future may reveal that anatoxin-a is more toxic than has been suggested in the currently available literature.

This recommended TDI should be considered provisional because of the paucity of toxicity data. OHA will update this TDI when more toxicity information becomes available.

Additional studies supporting this TDI: OHA only identified two primary studies that employed oral administration of anatoxin-a: the Fawell, et.al. study selected as the critical study (Fawell and James, 1994; Fawell et al., 1999b), and an older study conducted by Astrachan, et al. (Astrachan et al., 1980; Astrachan and Archer, 1981).

Independent reviews (Duy et al., 2000; Codd et al., 2005) of this Astrachan, et al. study have derived a TDI of 0.51 µg/kg-day, a value similar within a factor of 5 to the TDI selected (0.1 µg/kg-day). California's Environmental Protection Agency (CalEPA) has proposed an oral reference dose of 0.5 µg/kg-day (CalEPA, 2012), a value similar within a factor of 5 to the TDI selected here.

Other toxicity studies (Rogers et al., 2005) have been conducted using non-oral (mainly intraperitoneal injection) routes of exposure. Because human exposures to anatoxin-a in Oregon is expected to be primarily through ingestion, either in drinking water or accidental ingestion of surface water while recreating, OHA only considered studies using the oral route of exposure.

#### Provisional Recreational Water Guideline Value

OHA used the TDI of 0.1 µg/kg-day to derive a provisional recreational water **guideline value of 8 µg/L for anatoxin-A:**

$$\text{Guideline Value} = \frac{\text{TDI} \times \text{RSC} \times \text{BW}}{\text{IR}}$$

*Where:*

TDI = Tolerable Daily Intake (0.1 µg/kg-day)

RSC = 0.8 (U.S. EPA 2000a)

BW = Mean body weight of children 6 to < 11 years (31.8 kg) (U.S. EPA 2011)

IR = Recreational water incidental ingestion rate for children (0.33 L/d) at approximately the 90th percentile (U.S. EPA 2011; U.S. EPA 1997)

The GV for anatoxin-A was the result of new research on exposure factors provided by the Environmental Protection Agency (EPA) for microcystin and cylindrospermopsin, specifically affecting body weight and ingestion rate factors. These same factors were used to calculate the GV for anatoxin-A.

This recreational water guideline value is based on a provisional TDI. Therefore, this guideline value should also be considered provisional and subject to change should the provisional TDI be updated to accommodate new scientific information.

## Summary

OHA adopted health-based GVs for anatoxin-A:

- Tolerable Daily Intake: 0.1 µg/kg-day
- **Recreational Water Advisory Guideline Value: 8 µg/L**

As noted above, very few studies have been done to quantify the oral dose-response to anatoxin-a. Therefore, these GVs should be viewed as provisional and subject to revisions pending further research relevant to anatoxin-a toxicity.

## *Saxitoxins*

### Background

Saxitoxins (STXs) are a family of biological toxins associated with paralytic shellfish poisoning (PSP). This family includes saxitoxin (STX), neosaxitoxin (neoSTX), gonyautoxins, (GTX), C-toxins (C), 11-hydroxy-STX and decarbamoylsaxitoxins (dcSTXs)(van Apeldoorn et al., 2007). Because individual STXs vary in their toxicity, the European Food Safety Authority (EFSA) developed toxic equivalency factors (TEFs), based on toxicity in mice, so individual toxin concentrations can be considered relative to the toxicity of STX (EFSA, 2009). The proposed TEFs are: STX = 1, NeoSTX = 1, GTX1 = 1, GTX2 = 0.4, GTX3 = 0.6, GTX4 = 0.7, GTX5 = 0.1, GTX6 = 0.1, C2 = 0.1, C4 = 0.1, dc-STX = 1, dc-NeoSTX = 0.4, dc-GTX2 = 0.2, GTX3 = 0.4, and 11-hydroxy-STX = 0.3 (EFSA, 2009).

OHA adopted these TEFs as the method for reporting STX-equivalents (STX-eq) results for public health analysis in Oregon. Most labs report total saxitoxins, which is also acceptable. Previously few waterbody managers tested for this cyanotoxin because it was considered an insignificant threat in the Northwest. However from 2009 to 2011, 4 of 30 Washington State lakes sampled tested positive for saxitoxin (Hardy and Farrer, 2011).

Given the documented presence of saxitoxin in Washington, it was important to determine whether this cyanotoxin was also present in Oregon. Since development of GVs for saxitoxins in recreational waters by OHA, this toxin has been detected in Oregon waters. OHA asks water body managers to provide saxitoxin data when a waterbody contains taxa of cyanobacteria associated with this toxin.

EFSA established an acute RfD for STX-eq of 0.5 µg STX-eq/kg-day (EFSA, 2009). This acute RfD is based on available intoxication reports in humans across the European population. This acute RfD represents an estimated NOAEL.

### Tolerable Daily Intake

OHA used the RfD/NOAEL described above (0.5 µg/kg-day) to derive a provisional TDI of 0.05 µg/kg-day as follows:

$$TDI = \frac{NOAEL}{UF}$$

Where:

TDI = Tolerable Daily Intake (0.05 µg/kg-day)

NOAEL = No Observable Adverse Effect Level (0.5 µg/kg-day)

UF = Uncertainty Factors (10 for limitations in the database).

This TDI is based on an acute toxicity study, so it is only applicable to acute or short-term exposure scenarios. OHA applied a total uncertainty factor of 10 for database limitations<sup>2</sup>. This is the only study of its kind for saxitoxin and additional studies may find a lower RfD.

For humans, no uncertainty factor for interspecies variability was needed since the data were from human illnesses. OHA also did not apply an uncertainty factor for individual variability since the EFSA study covered the general population which included sensitive individuals.

#### Provisional Recreational Water Guideline Value

OHA used the TDI of 0.05 µg/kg-day to derive a provisional recreational water **guideline value of 4 µg/L for SXT-eq**:

$$\text{Guideline Value} = \frac{\text{TDI} \times \text{RSC} \times \text{BW}}{\text{IR}}$$

Where:

TDI= Acute oral reference dose (0.05 µg STX-eq/kg-day)

RSC = 0.8 (U.S. EPA 2000a)

BW = Mean body weight of children 6 to < 11 years (31.8 kg) (U.S. EPA 2011)

IR = Recreational water incidental ingestion rate for children (0.33 L/d) at approximately the 90th percentile (U.S. EPA 2011; U.S. EPA 1997)

The GV for saxitoxin was the result of new research on exposure factors provided by the Environmental Protection Agency (EPA) for microcystin and cylindrospermopsin, specifically affecting body weight and ingestion rate factors. These same factors were used to calculate the GV for saxitoxin.

OHA applies this SXT-eq guideline value to total saxitoxin results. This provisional recreational water guideline value is based on EFSA's acute RfD. This value is subject to change should additional toxicological information become available in the future.

#### Summary

OHA adopted a recreational water advisory guideline value of 4 µg STX-eq/L for saxitoxins. As noted above, this guideline value should be viewed as provisional and subject to revisions pending further research relevant to STX toxicity.

### ***Cylindrospermopsin***

#### Background

Previously, few waterbody managers tested for this cyanotoxin because it had been considered an insignificant threat in the Northwest. However, in 2011, a water body in Washington tested positive for cylindrospermopsin (Hardy and Farrer, 2011). Since 2011, cylindrospermopsin has

---

<sup>2</sup>OPHD did not originally apply the uncertainty factor for database limitations to the TDI for saxitoxins. Application of this uncertainty factor dropped OPHD's previous TDI and all GVs based on that TDI (recreational water and drinking water GVs) by a factor of 10. OPHD applied the database limitation uncertainty factor in this revision in keeping with the Ohio EPA, which first applied this uncertainty factor in 2014.

been detected in Oregon above the recreational guideline value established by OHA. Given the documented presence of cylindrospermopsin in Washington and Oregon, OHA asks waterbody managers to provide cylindrospermopsin data when a waterbody contains taxa of cyanobacteria associated with this toxin.

### Tolerable Daily Intake

To develop a TDI for cylindrospermopsin, OHA used the same study by Humpage et. al., 2003 that the EPA selected as the critical study in development of their 10-day Health Advisory for cylindrospermopsin. This 11-week study used male Swiss albino mice in which groups of mice were dosed with 0, 30, 60, 120, or 240 µg/kg-day (10 mice per dose group) of purified cylindrospermopsin by daily gavage. Authors monitored food and water consumption and body weights throughout the study. At nine weeks, authors conducted clinical exams with a focus on physiological and behavioral signs of toxicity. Near the end of the study an extensive panel of parameters was measured in serum and urine along with hematological endpoints. No deaths were reported in the study. Upon necropsy, organs were weighed and all tissues were examined histologically. The most sensitive endpoint observed was kidney weight, which increased in a dose-dependent manner starting at 60 µg/kg-day. The EPA selected 60 µg/kg-day from this study as the LOAEL and 30 µg/kg-day as the NOAEL [23].

Consistent with EPA's Health Advisory methodology, OHA applied a total uncertainty factor of 300 to the NOAEL of 30 µg/kg-day. The total UF of 300 was a composite of an UF of 10 for interspecies variability, 10 for individual variability, and 3<sup>3</sup> for database limitations. OHA used the NOAEL of 30 µg/kg-day to derive a provisional TDI of 0.1 µg/kg-day as follows:

$$\text{TDI} = \frac{\text{NOAEL}}{\text{UF}}$$

*Where:*

TDI = Tolerable Daily Intake (0.1 µg/kg-day)

NOAEL = No Observable Adverse Effect Level (30 µg/kg-day)

UF = Uncertainty Factors (300).

The EPA has also adopted this same TDI as their reference dose (RfD) for Cylindrospermopsin.

### Provisional Recreational Water Guideline Value

To derive a recreational water guideline value, OHA applied exposure factors to the TDI derived above (0.1 µg/kg-day) as follows:

$$\text{Guideline Value} = \frac{\text{TDI} \times \text{RSC} \times \text{BW}}{\text{IR}}$$

*Where:*

TDI = Oral reference dose (0.1 µg/kg-day)

RSC = 0.8 (U.S. EPA 2000a)

BW = Mean body weight of children 6 to < 11 years (31.8 kg) (U.S. EPA 2011)

---

<sup>3</sup> The previous assessment of cylindrospermopsin included a database limitation factor of 10. An uncertainty factor of 3 was used in the current 10-day Health Advisory issued by the EPA's Office of Water on June 17, 2015. To be consistent with EPA guidance, OPHD adopted this uncertainty factor which resulted in an increase in the TDI from the previous value by an approximate factor of 3.

IR = Recreational water incidental ingestion rate for children (0.33 L/d) at approximately the 90th percentile (U.S. EPA 2011; U.S. EPA 1997)

The mean body weight (BW) of 31.8 kg was used to represent a child between the age of 6 and 11 years. An incidental ingestion rate (IR) was based on EPA guidance for incidental ingestion of recreational water for children at the 90th percentile.

The GV for cylindrospermopsin was the result of new research on exposure factors provided by the Environmental Protection Agency (EPA), specifically affecting body weight and ingestion rate factors.

#### Summary

As noted above, OHA adopted 8 µg/L as a health-based guideline value for cylindrospermopsin in recreational water based on EPA criteria.

## Appendix C: Exposure pathways

The primary pathway for exposure to cyanotoxins is ingestion of water. Dermal effects are possible from the lipopolysaccharides found on cell surfaces, however cyanotoxins are not likely to cross the skin barrier and enter the bloodstream. Inhalation and aspiration of toxin is possible, especially through activities where the toxin is aerosolized, such as water skiing or splashing.

Ingestion of water can occur through both incidental and intentional ingestion. The risk of incidental ingestion is particularly high for children playing in near-shore areas where scum tends to accumulate. Exposure levels can be broadly defined as high, moderate and low based on recreational activity (Table C-1).

Table C-1. Level of recreational activity (modified from Queensland Health, 2001)

<i>Level of Exposure</i>	<i>Recreational Activity</i>
High	Swimming, diving, water skiing
Moderate	Canoeing, sailing, rowing
Low to none	Fishing, pleasure cruising, picnicking, hiking

Two possible scenarios for human intentional ingestion of recreational water should be considered. One is lake water used for drinking or cooking purposes by campers and hikers. Boiling, filtering or treating contaminated water with camping equipment will not make it potable. The second risk for exposure occurs when people draw in-home water directly from a lake or river. Private treatment systems have not proven effective in removing algae toxins. This exposure information is addressed in all advisory news releases, educational materials and signs.

### Public Drinking Water Systems

Drinking water is another exposure pathway of concern for cyanotoxins. Occasionally, cyanoHABs occur in waters that are drinking water sources. OHA's Drinking Water program has adopted the acute toxicity values for cyanotoxins in drinking water established by the EPA (Table C-2). Drinking water containing toxins above the acute values in Table C-2 could cause immediate harm to public health. Although these are not enforceable Maximum Contaminant Levels (MCLs), OHA recommends that public water systems use them as "Do Not Drink" thresholds.

For information regarding these guidelines, contact OHA at 971-673-0400 or [HAB.health@state.or.us](mailto:HAB.health@state.or.us). For more guidance specific to drinking water system operators, visit: <http://public.health.oregon.gov/HealthyEnvironments/DrinkingWater/Operations/Treatment/Pages/algae.aspx>.

Table C-2. Acute or short-term drinking water cyanotoxin toxicity values (µg/L)

<i>Drinking Water Guidance Value:</i>	<i>Microcystin</i>	<i>Cylindrospermopsin</i>	<i>Saxitoxin</i>	<i>Anatoxin-a</i>
Adults	1.6	3	1.6*	3
Ages 5 years and younger	0.3	0.7	0.3	0.7

Note: Rounding conventions are consistent with EPA's 10-day Health Advisories

\*OHA's previous drinking water guidance value for saxitoxin was 3 µg/L and was based on guidance used in other countries and not a TDI. This new drinking water value is based on the TDI established in Appendix B.

Table C-3 lists the exposure factors used to calculate drinking water GVs using the TDIs established in Appendix B. The equation used to calculate drinking water guidelines is identical to the equation used to calculate recreational GVs in Appendix B.

Table C-3. Exposure factors used to calculate drinking water GVs

Parameter	Adults	Children 5 and younger
Body Weight	80 kilograms	---
Intake Rate	2.5 liters	---
Body Weight-Normalized Intake Rate	---	0.15 liters/kilogram-body weight per day

Note: OHA adopted EPA’s exposure factors used in their derivation of 10-day Health Advisories for microcystin and cylindrospermopsin and applied them to the TDIs OHA derived for anatoxin-a and saxitoxins as well.

Fish Consumption

At this time, there is insufficient information to determine the risk of consuming fish caught in waters with a cyanoHAB. Studies have shown that toxins mainly accumulate in the liver and viscera of fish, and microcystin has been detected in the fillet (Vasconcelos, 1999; de Magalhaes et al., 2001; Kann, 2008; Washington Department of Ecology, 2010; Kann et al., 2011). At a minimum, organs and skin should be removed and discarded prior to cooking fillets and caution should be taken with shellfish as cyanotoxins have been shown to accumulate in edible tissue (Vasconcelos, 1999).

Risk to Animals

Animals are extremely sensitive to cyanotoxins. Routes of exposure are ingestion when pets and wildlife drink water from a harmful algae-filled lake or pond, lick their fur after swimming or eat dried cells that accumulate along the shoreline. If toxins are present when animals drink the water, the animals can become very ill and possibly die.

Because dogs are cyanotoxin sensitive animals and dog deaths have been confirmed due to cyanoHABs, OHA developed dog-specific GVs for cyanotoxins in recreational water (Table C-4).

Table C-4. Dog-specific GVs for cyanotoxins in recreational waters (µg/L)

<i>Dog Guidance Value:</i>	<i>Anatoxin-a</i>	<i>Cylindrospermopsin</i>	<i>Microcystin</i>	<i>Saxitoxin</i>
	0.4	0.4	0.2	0.02

Note: All dog-specific GVs have been changed in this revision because California EPA’s estimate of the amount of water an exercising dog consumes per kilogram body weight was updated in 2012 (from 0.168 to 0.255 L/kg-day). Current dog-specific GVs are now consistent with the

California EPA update. The dog-specific value for saxitoxins was further modified by application of an uncertainty factor to the dog-specific TDI for interspecies differences in sensitivity between humans (the species in the critical study) and dogs.

OHA does not intend to use these dog-specific guidelines values as the basis for public health advisories. Rather, they are offered as a resource to veterinarians and veterinary associations to use as appropriate. OHA will use these values and potential exposure scenarios in discussions with individual veterinarians or pet owners to educate them on the vulnerability of pets to cyanotoxin exposures. Contact OHA for details about the origin of these dog-specific values.

## Appendix D: References

van Apeldoorn, M.E., van Egmond, H.P., Speijers, G.J.A., and Bakker, G.J.I. (2007). Toxins of cyanobacteria. *Molecular Nutrition & Food Research* 51, 7–60.

Astrachan, N.B., and Archer, B.G. (1981). Simplified monitoring of anatoxin-a by reverse-phase high performance liquid chromatography and the sub-acute effects of anatoxin-a in rats. In *The Water Environment: Algal Toxins and Health*, W.W. Carmichael, ed. (New York, NY: Plenum Press), pp. 437–446.

Astrachan, N.B., Archer, B.G., and Hilbelink, D.R. (1980). Evaluation of the subacute toxicity and teratogenicity of anatoxin-a. *Toxicon : Official Journal of the International Society on Toxinology* 18, 684–688.

ATSDR (2005). *Public Health Assessment Guidance Manual* (Atlanta, GA: US Department of Health and Human Services).

Burch, M.D. (2008). Effective doses, guidelines & regulations. *Advances in Experimental Medicine and Biology* 619, 831–853.

CalEPA (2012). *Toxicological Summary and Suggested Action Levels to Reduce Potential Adverse Health Effects of Six Cyanotoxins* (Sacramento, CA: California Environmental Protection Agency).

Carey, C.C., Haney, J.F., and Cottingham, K.L. (2007). First report of microcystin-LR in the cyanobacterium *Gloeotrichia echinulata*. *Environmental Toxicology* 22, 337–339.

Chorus, I., and Bartram, J. (1999). *Toxic cyanobacteria in water: a guide to their public health consequences, monitoring and management*. WHO.

Chorus, I., Falconer, I.R., Salas, H.J., and Bartram, J. (2000). HEALTH RISKS CAUSED BY FRESHWATER CYANOBACTERIA IN RECREATIONAL WATERS. *Journal of Toxicology and Environmental Health, Part B* 3, 323–347.

Codd, G.A., Morrison, L.F., and Metcalf, J.S. (2005). Cyanobacterial toxins: risk management for health protection. *Toxicol Appl Pharmacol* 203, 264–272.

Dang, W. (1996). *The Swimmer Exposure Assessment Model (SWIMODEL) and its use in estimating risks of chemical use in swimming pools* (US Environmental Protection Agency).

Duy, T.N., Lam, P.K., Shaw, G.R., and Connell, D.W. (2000). Toxicology and risk assessment of freshwater cyanobacterial (blue-green algal) toxins in water. *Reviews of Environmental Contamination and Toxicology* 163, 113–185.

EFSA (2009). *Scientific Opinion: Marine biotoxins in shellfish -- Saxitoxin group*. *The EFSA Journal* 1019, 1–76.

Falconer, I.R., and Buckley, T.H. (1989). Tumour promotion by *Microcystis* sp., a blue-green alga occurring in water supplies. *Medical Journal of Australia* 150, 351–352.

Falconer, I.R., and Humpage, A.R. (2005). Health risk assessment of cyanobacterial (blue-green algal) toxins in drinking water. *International Journal of Environmental Research and Public Health* 2, 43–50.

Fawell, J.F., and James, H.A. (1994). Toxins from blue-green algae: Toxicological assessment of anatoxin-a and a method for its determination in reservoir water.

Fawell, J.K., Mitchell, R.E., Everett, D.J., and Hill, R.E. (1999a). The Toxicity of Cyanobacterial Toxins in the Mouse: I microcystin-LR. *Hum Exp Toxicol* 18, 162–167.

Fawell, J.K., Mitchell, R.E., Hill, R.E., and Everett, D.J. (1999b). The toxicity of cyanobacterial toxins in the mouse: II anatoxin-a. *Human & Experimental Toxicology* 18, 168–173.

Funari, E., and Testai, E. (2008). Human health risk assessment related to cyanotoxins exposure. *Crit Rev Toxicol* 38, 97–125.

Hardy, J., and Farrer, D. (2011). Personal communication.

Heinze, R. (1999). Toxicity of the cyanobacterial toxin microcystin-LR to rats after 28 days intake with the drinking water. *Environmental Toxicology* 14, 57–60.

Humpage, A.R., Falconer, I.R. (2003) Oral toxicity of the cyanobacterial toxin cylindrospermopsin in male swiss albino mice: Determination of no observable adverse effect level for deriving a drinking water guideline value. *Environmental Toxicology* 18, 94-103.

Kann, J. (2008). Microcystin Bioaccumulation in Klamath River Fish and Freshwater Mussel Tissue: Preliminary 2007 Results (Ashland, OR: Aquatic Ecosystem Sciences, LLC.).

Kann, J., Bowater, L., Johnson, G., and Bowman, C. (2011). Preliminary 2010 Microcystin Bioaccumulation Results for Klamath River Salmonids (Updated 4-7-2011). (Ashland, OR: Aquatic Ecosystem Sciences, LLC. Karuk Tribe Department of Natural Resources).

Kann, J., and Corum, S. (2009). Toxigenic *Microcystis aeruginosa* bloom dynamics and cell density/chlorophyll a relationships with microcystin toxin in the Klamath River, 2005-2008 (Karuk Tribe Department of Natural Resources).

de Magalhaes, V.F., Soares, R.M., and Azevedo, S.M.F.O. (2001). Microcystin contamination in fish from the Jacarepagua Lagoon (Rio de Janeiro, Brazil): ecological implication and human health risk. *Toxicon* 39, 1077–1085.

Manganelli, M., Scardala, S., Stefanelli, M., Vichi, S., Mattei, D., Bogialli, S., Ceccarelli, P., Corradetti, E., Petrucci, I., Gemma, S., et al. (2010). Health risk evaluation associated to *Planktothrix rubescens*: An integrated approach to design tailored monitoring programs for human exposure to cyanotoxins. *Water Research* 44, 1297–1306.

New Zealand Ministry of Health (2002). Cyanobacteria Data Sheet.

Ohio EPA (2014). Public Water System Harmful Algal Bloom Response Strategy. Ohio EPA, Columbus, Ohio.

Pegram, R.A., Nichols, T., Etheridge, S., Humpage, A., LeBlanc, S., Love, A., Neilan, B., Pflugmacher, S., Runnegar, M., and Thacker, R. (2008). Cyanotoxins Workgroup report. *Advances in Experimental Medicine and Biology* 619, 317–381.

Pilotto, L., Hobson, P., Burch, M.D., Ranmuthugala, G., Attewell, R., and Weightman, W. (2004). Acute skin irritant effects of cyanobacteria (blue-green algae) in healthy volunteers. *Australian and New Zealand Journal of Public Health* 28, 220–224.

Queensland Health (2001). *Cyanobacteria in Recreational and Drinking Waters*. Environmental Health Assessment Guidelines (Environmental Health Unit).

Rinehart, K., Namikoshi, M., and Choi, B. (1994). Structure and biosynthesis of toxins from blue-green algae (cyanobacteria). *Journal of Applied Phycology* 6, 159–176.

Rogers, E.H., Hunter, E.S., Moser, V.C., Phillips, P.M., Herkovits, J., Muñoz, L., Hall, L.L., and Chernoff, N. (2005). Potential developmental toxicity of anatoxin-a, a cyanobacterial toxin. *Journal of Applied Toxicology : JAT* 25, 527–534.

Short, S.B., and Edwards, W.C. (1990). Blue-green algae toxicoses in Oklahoma. *Veterinary and Human Toxicology* 32, 558–560.

Turner, P.C., Gammie, A.J., Hollinrake, K., and Codd, G.A. (1990). Pneumonia associated with contact with cyanobacteria. *BMJ* 300, 1440–1441.

Ueno, Y., Makita, Y., Nagata, S., Tsutsumi, T., Yoshida, F., Tamura, S.-I., Sekijima, M., Tashiro, F., Harada, T., and Yoshida, T. (1999). No chronic oral toxicity of a low dose of microcystin-LR, a cyanobacterial hepatotoxin, in female BALB/c mice. *Environmental Toxicology* 14, 45–55.

USEPA (2006). *Toxicological Reviews of Cyanobacterial Toxins: Anatoxin-a* (External Review Draft) (US Environmental Protection Agency).

USEPA (2006b). *Toxicological Reviews of Cyanobacterial Toxins: Cylindrospermopsin* (External Review Draft) (US Environmental Protection Agency).

USEPA (2014). *Policies and Guidelines*. Available online: <http://www2.epa.gov/nutrient-policy-data/policies-and-guidelines> (accessed on 13 November 2014).

Vasconcelos, V.M. (1999). Cyanobacterial toxins in Portugal: effects on aquatic animals and risk for human health. *Brazilian Journal of Medical and Biological Research* 32, 249–254.

Voloshko, L.N., Plyushch, A.V., and Titova, N.N. (2008). Toxins of Cyanobacteria (Cyanophyta). *International Journal on Algae* 10, 14–33.

Washington Department of Ecology (2010). *Blue-Green Algae Toxins in Washington Lakes: Screening Fish Tissues for Microcystins and Anatoxin-a* (Ecology).

Washington Department of Health (2008). *Washington State Recreational Guidance for Microcystins (Provisional) and Anatoxin-a (Interim/Provisional)* (Olympia, WA: Washington Department of Health).

Washington Department of Health (2011). *Washington State Recreational Guidance for Cylindrospermopsin (Provisional) and Saxitoxin (Provisional)* (Health).